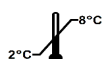


Instructions for use
Estradiol rat ELISA

Please use only the valid version of the Instructions for Use provided with the kit

REF**AR E-8800R**

96

RUO

For research
use only –
Not for use
in diagnostic
procedures

Estradiol rat ELISA

1. INTRODUCTION

1.1 Intended use

The **Estradiol rat ELISA** is a competitive immunoassay for the quantitative measurement of estradiol in rat serum. **For research use only – Not for use in diagnostic procedures.**

1.2 Summary and explanation

Estradiol (E2 or 17 β -estradiol) is an estrogenic hormone produced by the ovaries and in smaller amounts by the adrenal cortex and testes. It is the most potent female sexual hormone and is essential for maintaining normal female functions. During the oestrous cycle which can be divided into four phases (proestrus, estrus, metestrus, diestrus) estradiol concentrations rise gradually from metestrus to proestrus and fall to barely detectable levels in estrus. The maximum concentration is reached around mid-day of proestrus (1, 2). Apart from its effects on sexual characteristics it has important influence on the growth and development of the brain (3, 4).

In female rodents, the determination of estradiol is a useful marker in evaluating and monitoring the state of the reproductive functions and pregnancy as well.

2. PRINCIPLE

The Estradiol rat ELISA Kit is a solid phase enzyme-linked immunosorbent assay (ELISA), based on the principle of competitive binding.

The microtiter wells are coated with an anti-Estradiol antibody. An unknown amount of estradiol present in the sample competes with an Estradiol-horseradish peroxidase conjugate for binding to the coated antibody. After incubation the unbound conjugate is washed off. The amount of bound peroxidase conjugate is inversely proportional to the concentration of Estradiol in the sample. After addition of the substrate solution, the intensity of color developed is inversely proportional to the concentration of free Estradiol in the sample.

3. WARNINGS AND PRECAUTIONS

1. This kit is intended for research only. Use by staff, who is specially informed and trained in methods which are carried out by use of immunoassays.
2. All blood components and biological materials should be handled as potentially hazardous in use and for disposal. Follow universal precautions when handling and disposing of infectious agents.
3. Before starting the assay, read the instructions completely and carefully. Use the valid version of the package insert provided with the kit. Be sure that everything is understood.
4. The microplate contains snap-off strips. Unused wells must be stored at 2 – 8 °C in the sealed foil pouch and used in the frame provided.
5. Pipetting of samples and reagents must be done as quickly as possible and in the same sequence for each step.
6. Use reservoirs only for single reagents. This especially applies to the substrate reservoirs. Using a reservoir for dispensing a substrate solution that had previously been used for the conjugate solution may turn solution colored. Do not pour reagents back into vials as reagent contamination may occur.
7. Mix the contents of the microplate wells thoroughly to ensure good test results. Do not reuse microwells.
8. Do not let wells dry during assay; add reagents immediately after completing the rinsing steps.
9. Allow the reagents to reach room temperature (18 – 25 °C) before starting the test. Temperature will affect the absorbance readings of the assay.
10. Never pipet by mouth and avoid contact of reagents and samples with skin and mucous membranes.
11. Do not smoke, eat, drink or apply cosmetics in areas where samples or kit reagents are handled.
12. Wear disposable latex gloves when handling samples and reagents. Microbial contamination of reagents or samples may give false results.
13. Handling should be done in accordance with the procedures defined by an appropriate national biohazard safety guideline or regulation.
14. Do not use reagents beyond expiry date as shown on the kit labels.
15. All indicated volumes have to be performed according to the protocol. Optimal test results are only obtained when using calibrated pipettes and microtiterplate readers.
16. Do not mix or use components from kits with different lot numbers. It is advised not to exchange wells of different plates even of the same lot. The kits may have been shipped or stored under different conditions and the binding characteristics of the plates may result slightly different.
17. Avoid contact with Stop Solution. It may cause skin irritation and burns.
18. Some reagents contain Proclin 300, CMIT and/or MIT as preservatives. In case of contact with eyes or skin, flush immediately with water.
19. Chemicals and prepared or used reagents have to be treated as hazardous waste according to the national biohazard safety guideline or regulation.
20. For information please refer to Safety Data Sheets. Safety Data Sheets for this product are available upon request directly from the manufacturer.

21. If product information, including labeling, is incorrect or inaccurate, please contact the kit manufacturer or supplier.







4. REAGENTS

4.1 Reagents provided

AR E-8831 **Microtiter Plate**

Content: 12 x 8 (break apart) strips with 96 wells; wells coated with anti-Estradiol antibody

Standards – Lyophilized; For reconstitution see "Reagent Preparation"

Cat. no.	Symbol	Standard	Concentration
AR E-8801		Standard A	0 pg/ml
AR E-8802		Standard B	5 pg/ml
AR E-8803		Standard C	20 pg/ml
AR E-8804		Standard D	80 pg/ml
AR E-8805		Standard E	320 pg/ml
AR E-8806		Standard F	1280 pg/ml

AR E-8813 **Incubation Buffer** – Ready to use

Volume: 1 x 7 ml

AR E-8841 **Enzyme Conjugate Diluent** – Ready to use

Volume: 1 x 30 ml

Hazards identification:



H317 May cause an allergic skin reaction.

AR E-8840 **Enzyme Conjugate** – 100X concentrated

Content: 17β-Estradiol labeled horseradish peroxidase in buffered matrix

Volume: 1 x 0.3 ml;
See "Reagent Preparation" (4.4)

Hazards identification:



H317 May cause an allergic skin reaction.

AR E-0055 **Substrate Solution** – Ready to use

Content: Contains tetramethylbenzidine (TMB) and hydrogen peroxide in a buffered matrix.

Volume: 1 x 22 ml

AR E-0080 **Stop Solution** – Ready to use

Content: Contains 2 N Hydrochloric Acid solution.

Volume: 1 x 7 ml

Hazards identification:



H290 May be corrosive to metals.
H314 Cause severe skin burns and eye damage.
H335 May cause respiratory irritation.

AR E-0030 **Wash Solution** – 10X concentrated

Volume: 1 x 50 ml;
See "Reagent Preparation" (4.4)

4.2 Materials required but not provided

- Microcentrifuge
- A microtiter plate reader capable for endpoint measurement at 450 nm
- Calibrated variable precision micropipettes (50 µl, 75 µl, 200 µl, 300 µl).
- Microplate mixer operating at 900 rpm
- Vortex mixer
- Absorbent paper
- Distilled or deionized water
- Timer
- Semi logarithmic graph paper or software for data reduction

4.3 Storage conditions

When stored at 2 °C to 8 °C unopened reagents will be stable until expiration date. Do not use reagents beyond this date. Opened reagents must be stored at 2 – 8 °C. After first opening the reagents are stable for 30 days if used and stored properly. Microtiter wells must be stored at 2 °C to 8 °C. Take care that the foil bag is sealed tightly. Protect TMB-Substrate Solution from light.

Store Standards refrigerated, they will be stable at 2 – 8 °C for 7 days after reconstitution. For longer storage freeze at -20 °C.

4.4 Reagent preparation

Allow the reagents and the required number of wells to reach room temperature (18 – 25 °C) before starting the test.

Wash Solution:

Dilute 50 ml of 10X concentrated Wash Solution with 450 ml deionized water to a final volume of 500 ml. The diluted Wash Solution is stable for at least 3 months at room temperature (18 – 25 °C). Precipitates may form when stored at 2 – 8 °C, which should dissolve again by swirling at room temperature (18 – 25 °C). The wash solution should only be used when the precipitates have completely dissolved.

Standards:

Reconstitute lyophilized standards with **0.5 ml deionized water** 30 min. before use.

Enzyme Conjugate:

Immediately before use dilute Enzyme Conjugate Concentrate 1:100 in Enzyme Conjugate Diluent, for example 0.1 ml concentrated Enzyme Conjugate Concentrate + 9.9 ml Enzyme Conjugate Diluent. Mix thoroughly.

Attention: Please prepare only immediately before use!

4.5 Disposal of the kits

The disposal of the kit must be made according to the national regulations. Special information for this product is given in the Safety Data Sheet.

4.6 Damaged test kits

In case of any severe damage of the test kit or components, the manufacturer have to be informed written, latest one week after receiving the kit. Severely damaged single components should not be used for a test run. They have to be stored until a final solution has been found. After this, they should be disposed according to the official regulations.

5. SAMPLE COLLECTION AND STORAGE

For determination of rat estradiol **serum samples** can be used. The procedure calls for 75 µl sample per well. The samples should assay immediately or aliquot and stored at -20 °C. Avoid repeated freeze-thaw cycles. Samples expected to contain estradiol concentrations higher than the highest standard (1280 pg/ml) should be diluted with the zero standard before assay. The additional dilution step has to be taken into account for the calculation of the results.

6. ASSAY PROCEDURE

6.1 General Remarks

- All reagents and samples must be allowed to come to room temperature before use. All reagents must be mixed without foaming.
- Once the test has been started, all steps should be completed without interruption.
- Use new disposal plastic pipette tips for each standard, control or sample in order to avoid cross contamination.
- Absorbance is a function of the incubation time and temperature. Before starting the assay, it is recommended that all reagents are ready, caps removed, all needed wells secured in holder, etc. This will ensure equal elapsed time for each pipetting step without interruption.
- As a general rule the enzymatic reaction is linearly proportional to time and temperature.

- Respect the incubation times as stated in this instructions for use.
- Duplicate determination of standards, controls and samples is recommended in order to identify potential pipetting errors.
- A standard curve must be established for every run.
- Microtiter plate washing is important. Improperly washed wells will give erroneous results. It is recommended to use a multichannel pipette or a multistepper, respectively, or an automatic microtiter plate washing system. Do not allow the wells to dry between incubations. Do not scratch coated wells during rinsing and aspiration. Rinse and fill all reagents with care. While rinsing, check that all wells are filled precisely with wash solution, and that there are no residues in the wells.
- For internal quality control we suggest to use **RAT CONTROL SET coded AR K-8000R. For more information please contact the manufacturer.**

6.2 Assay Procedure

1.	Prepare a sufficient number of microplate wells to accommodate standards, controls and samples in duplicates.
2.	Dispense 75 µl of each Standard, Controls and Sample <u>with new disposable tips</u> into appropriate wells.
3.	Dispense 50 µl of Incubation Buffer into each well.
4.	Incubate for 120 minutes at room temperature (18 – 25 °C) on a plate shaker (900 rpm).
5.	Immediately before use dilute the Enzyme Conjugate Concentrate 1:100 in Enzyme Conjugate Diluent (see chapter 4.4). Add 50 µl diluted Enzyme Conjugate into each well.
6.	Incubate for 60 minutes at room temperature (18 – 25 °C) on a plate shaker (900 rpm). Important Note: Optimal reaction in this assay is markedly dependent on shaking of the microplate!
7.	Discard the content of the wells and rinse the wells 4 times with diluted Wash Solution (300 µl per well). Remove as much Wash Solution as possible by beating the microplate on absorbent paper.
8.	Add 200 µl of Substrate Solution to each well.
9.	Incubate without shaking for 30 minutes in the dark.
10.	Stop the reaction by adding 50 µl of Stop Solution to each well.
11.	Determine the absorbance of each well at 450 nm. It is recommended to read the wells within 15 minutes.

6.3 Calculation of results

1. Calculate the average absorbance values for each set of standards, controls and samples.
2. The obtained optical density of the standards (y-axis, linear) are plotted against their corresponding concentrations (x-axis, logarithmic) either on semilogarithmic paper or using an automated method.
3. Using the mean absorbance value for each sample, determine the corresponding concentration from the standard curve.
4. Automated method: The results in the package insert have been calculated automatically using a 4 PL (4 Parameter Logistics) curve fit. 4 Parameter Logistics is the preferred calculation method. Other data reduction functions may give slightly different results.
5. The concentration of the samples can be determined directly from this standard curve. Samples with concentrations higher than that of the highest standard have to be further diluted. For the calculation of the concentrations, this dilution factor has to be taken into account.

Example of typical standard curve

Following data are intended for illustration only and should not be used to calculate results from another run.

Standard	Optical Units (450nm)
Standard A (0 pg/ml)	2.945
Standard B (5 pg/ml)	2.667
Standard C (20 pg/ml)	2.114
Standard D (80 pg/ml)	0.989
Standard E (320 pg/ml)	0.311
Standard F (1280 pg/ml)	0.129

7. EXPECTED VALUES

Each laboratory should establish its own normal ranges.

8. QUALITY CONTROL

Good laboratory practice requires that controls are run with each standard curve. A statistically significant number of controls should be assayed to establish mean values and acceptable ranges to assure proper performance. The use of control samples is advised to assure the day-to-day validity of results.

For internal quality control we suggest to use **RAT CONTROL SET coded AR K-8000R**.

Employ appropriate statistical methods for analysing control values and trends. If the results of the assay do not fit to the established acceptable ranges of control materials results should be considered invalid. In this case, please check the following technical areas: Pipetting and timing devices, microtiter plate reader, expiration dates of reagents, storage and incubation conditions, aspiration and washing methods. After checking the above mentioned items without finding any error contact your distributor or the manufacturer directly.

9. PERFORMANCE CHARACTERISTICS

9.1 Analytical sensitivity

The lowest analytical detectable level of Estradiol that can be distinguished from the Zero Standard is 2.5 pg/ml at the 2SD confidence limit.

9.2 Specificity (Cross Reactivity)

The following materials have been evaluated for cross reactivity. The percentage indicates cross reactivity at 50% displacement compared to Estradiol.

Steroid	% Cross reaction
Androstenedione	< 0.1
17-Hydroxyprogesterone	< 0.1
Corticosterone	< 0.1
Estriol	0.4
Estrone	4.2
Pregnenolone	< 0.1
E2-3-Glucuronide	3.8
E2-3-Sulphate	3.6
E2-17-Glucuronide	< 0.1
Progesterone	< 0.1
Testosterone	< 0.1
Fulvestrant	9.5

9.3 Assay dynamic range

The range of the assay is between 2.5 – 1280 pg/ml.

9.4 Reproducibility

9.4.1 Intra-Assay

The intra-assay variation was determined by 20 replicate measurements of three serum samples within one run. The within-assay variability is shown below:

	Serum 1	Serum 2	Serum 3
Mean (pg/ml)	29.9	118.9	246.3
SD	1.8	3.6	7.8
CV (%)	6.1	3.0	3.2
n =	20	20	20

9.4.2 Inter-Assay

The inter-assay (between-run) variation was determined by duplicate measurements of three serum samples in 11 different tests.

	Serum 1	Serum 2	Serum 3
Mean (pg/ml)	23.6	104.7	226.8
SD	1.6	4.1	16.0
CV (%)	7.0	4.0	7.1
n =	11	11	11

9.5 Recovery

Using the Standard Matrix two spiking solutions of 100 ng/ml and 10 ng/ml were prepared. 1000 µl of one serum was spiked with 2, 3.5 and 5 µl of the 100 ng/ml solution, and 1000 µl of two sera spiked with 5, 10 and 15 µl of the 10 ng/ml solution, leaving the serum matrices intact. All samples were measured by the Estradiol rat ELISA procedure.

Sample	Spiking (pg/ml)	Measured (pg/ml)	Expected (pg/ml)	Recovery (%)
1	–	20.7	–	–
	200	211.1	220.7	96%
	350	298.7	370.7	81%
	500	444.8	520.7	85%
2	–	19.4	–	–
	50	68.6	69.4	99%
	100	111.9	119.4	94%
	150	143.4	169.4	85%
3	–	11.2	–	–
	50	71.6	61.2	117%
	100	128.4	111.2	115%
	150	156.3	161.2	97%

9.6 Linearity

Three serum samples were assayed undiluted and diluted with the Standard A.

Serum	Dilution	Measured (pg/ml)	Expected (pg/ml)	Linearity (%)
1	–	196,7	./.	./.
	1 in 2	80,3	98,4	82%
	1 in 4	44,1	49,2	90%
	1 in 8	25,7	24,6	104%
2	–	231,3	./.	./.
	1 in 2	124,2	115,7	107%
	1 in 4	67,6	57,8	117%
	1 in 8	33,6	28,9	116%
3	–	359,9	./.	./.
	1 in 2	182,1	180,0	101%
	1 in 4	95,4	90,0	106%
	1 in 8	49,4	50,0	99%

10. LIMITATIONS OF PROCEDURE

Reliable and reproducible results will be obtained when the assay procedure is performed with a complete understanding of the package insert instruction and with adherence to good laboratory practice. Any improper handling of samples or modification of this test might influence the results.

10.1 Interfering Substances

- Do not use any hemolytic, icteric or lipemic samples to avoid any interferences.
- Samples containing sodium azide should not be used in the assay.
- Non-specific interferences with this in vitro immunoassay cannot be excluded. If unplausible results are suspected, they should be considered invalid and verified by further testing.

10.2 Drug Interferences

Any medication containing Estradiol will significantly influence the measurement of this analyte. The Estradiol ELISA should not be used for subjects being treated with the drug fulvestrant (Faslodex®) which cross reacts in the Estradiol ELISA and could lead to falsely elevated test results.

11. LEGAL ASPECTS

11.1 Reliability of Results

The test must be performed exactly as per the manufacturer's instructions for use. Moreover the user must strictly adhere to the rules of GLP (Good Laboratory Practice) or other applicable national standards and/or laws. This is especially relevant for the use of control reagents. It is important to always include, within the test procedure, a sufficient number of controls for validating the accuracy and precision of the test.

The test results are valid only if all controls are within the specified ranges and if all other test parameters are also within the given assay specifications. In case of any doubt or concern please contact the manufacturer.

11.2 Liability

Any modification of the test kit and/or exchange or mixture of any components of different lots from one test kit to another could negatively affect the intended results and validity of the overall test. Such modification and/or exchanges invalidate any claim for replacement.

Regardless, in the event of any claim, the manufacturer's liability is not to exceed the value of the test kit. Any damage caused to the test kit during transportation is not subject to the liability of the manufacturer.

12. REVISION HISTORY OF INSTRUCTION FOR USE

Changes from the previous version 4.0e to actual version 5.0

Chap. 3: updated

Chap. 4.2: updated (shaker needed at 900 rpm)

Chap. 6.1: additions (notes on the washing procedure)

Chap. 6.2: shaking speed at 900 rpm (previously > 600 rpm)

Chap. 8: added

Chap. 10: updated













Room temperature 18 – 25 °C (previously 21 – 26 °C)

Editorial changes

13. REFERENCES

1. Abubakar A. Shaikh
Estrone and estradiol levels in the ovarian venous blood from rat during the estrous cycle and pregnancy; Biology of Reproduction (1971); 5, 297 – 307.
2. Hawkins, R.A, Freedman, A., Marshall, A., Killen, E.
Oestradiol-17 β and prolactin levels in rat peripheral plasma; Br. J. Cancer (1975); 32, 179.
3. Zamani, M.R, Desmond, N.L, Levy, W.B..
Estradiol modulates long term synaptic depression in female rat hippocampus.
J. Neurophysiol (2000); 84: 1800 – 1808.
4. McCarthy, M.M.
Estradiol and the developing brain; Physiol. Rev. (2008); 88: 91 – 134.

Symbols:

	Storage temperature		Manufacturer		Contains sufficient for <n> tests
	Use-by date		Batch code		
	Consult instructions for use		Content		
	Caution		Catalogue number		Distributor
	Date of manufacture				For research use only!